

PCMD MicroCT Imaging Core Learning Lunch Series

Innovations in Bone Marrow Adipose Tissue Imaging **Optimizing Computational Workflows** **Introducing the New DXA Scanner**

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PCMD MicroCT Imaging Core

Penn Center for Musculoskeletal Disorders (P30-AR069619)

Outlines

- Brief introduction of our core facility
- Bone Marrow Adipose Tissue Imaging
- Optimizing Computational Workflows
- Q & A



µCT Imaging Core Resources

Model	Location	Scan Size (ØxL)	Voxel Size (µm)	Applications
µCT 35	Stemmler 335A	37.9 x 120	3.5-72	High resolution <i>ex vivo</i> scans
µCT 45	Stemmler 335A	50 x 120	3.0-100	High resolution <i>ex vivo</i> scans
vivaCT 80	Stemmler 368B	80 x 145	10.4-76	High resolution <i>in vivo</i> scans
New! DXA	Stemmler 368C	165 x 255	100	Full body 2D scan and analysis
µCT 50	PVAMC/TMRC	50 x 120	0.5-100	Ultra high resolution (sub-micron) <i>ex vivo</i> scans
vivaCT 75	PVAMC/TMRC	79.9 x 145	21-150	<i>In vivo</i> scans for small animals; <i>Ex vivo</i> scans for large specimens
XtremeCT II	CTRC	140 x 200	60-82	Clinical scans for peripheral skeleton



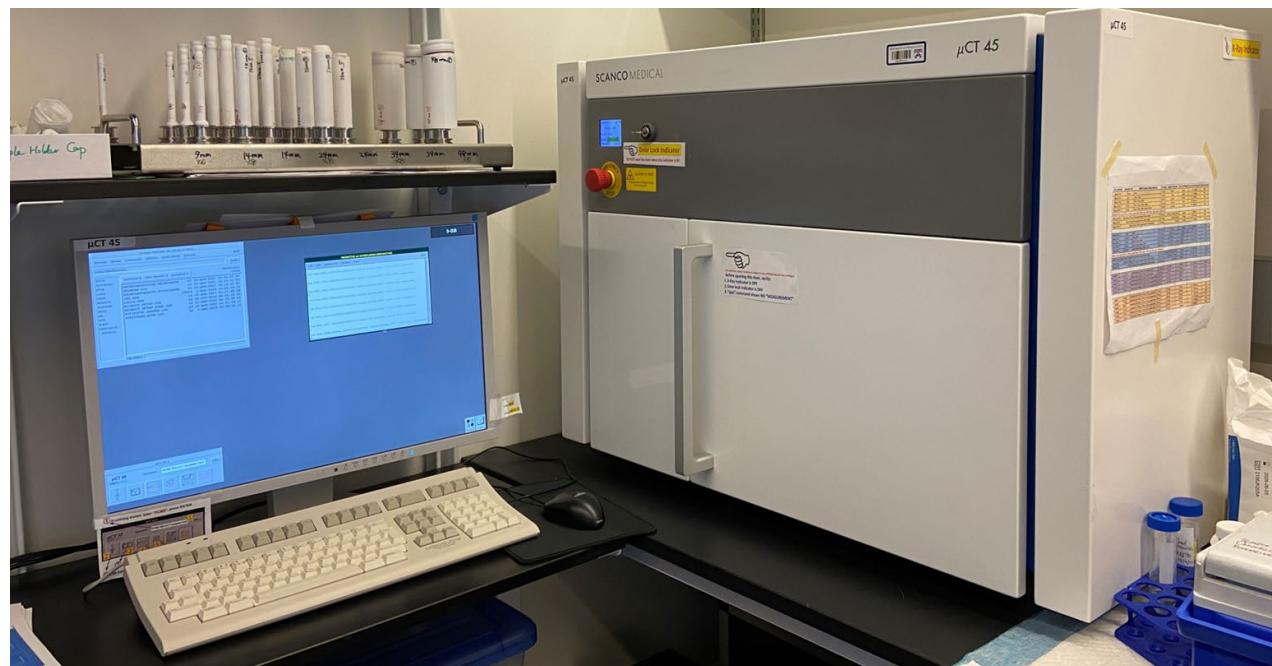
Ex vivo (Specimen) Scanners

- Scanco μ CT 35 (Purchased in 2012)
 - Native voxel sizes: 3.5 μ m, 6 μ m, 10 μ m, 15 μ m, 18.5 μ m



Ex vivo (Specimen) Scanners

- Scanco μ CT 45 (Purchased in 2019)
 - Native voxel sizes: 3 μ m, 4.5 μ m, 7.4 μ m, 10.4 μ m, 14.6 μ m
 - Carousel system supporting 20 sample holders
 - “Air” filter for scanning low density materials
 - “Copper” filter for scanning specimen with metal implants



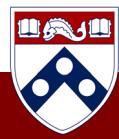
In vivo Scanner

- Scanco vivaCT 80 (Purchased in 2018)
 - Voxel sizes: 10.4 μm , 11.6 μm , 13 μm , 16.1 μm , 20.8 μm , 26 μm
 - Internal heating device to keep animal warm
 - Internal camera to monitor animal's breathing
 - Ex vivo scan for specimen from large animals or human cadaver



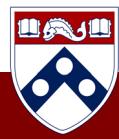
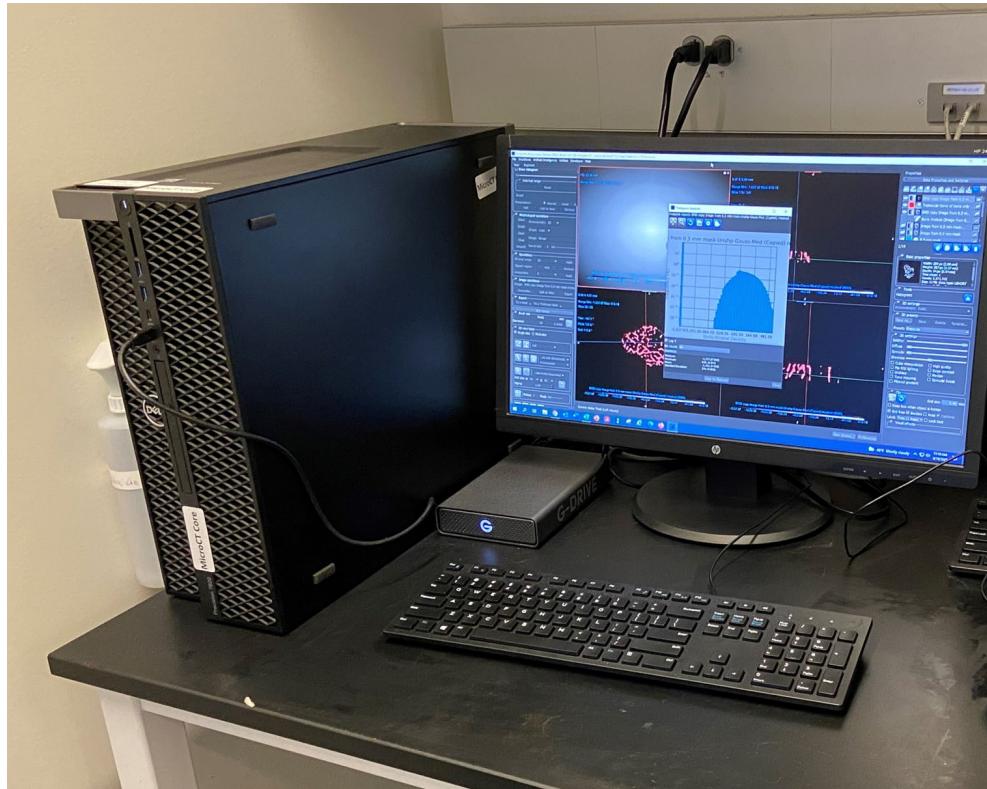
MicroCT Analysis PC

- 2 PCs for MicroCT Analysis (315 Stemmler)
 - Windows 10 platform
 - Either remote or onsite access
 - Scanco software



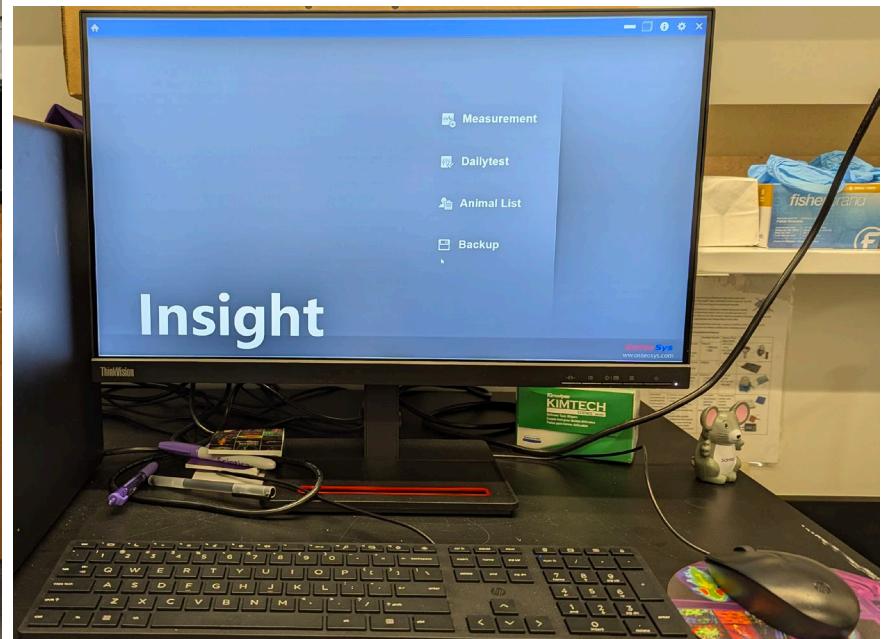
Dragonfly Workstation

- Workstation for Dragonfly software (324 Stemmler)
 - Windows 10 platform
 - PMACS account required (either remote or onsite access)
 - Deep learning assisted analysis
 - Training videos <https://www.theobjects.com/dragonfly/tutorials.html>



Scintica Insight DXA

- DXA system (368B Stemmler)
 - Windows 10 platform
 - In vivo / Ex vivo 2D scan and analysis
 - 25 sec per scan
 - Video tutorial: <https://www.youtube.com/watch?v=ZRWOm7NnK1g>



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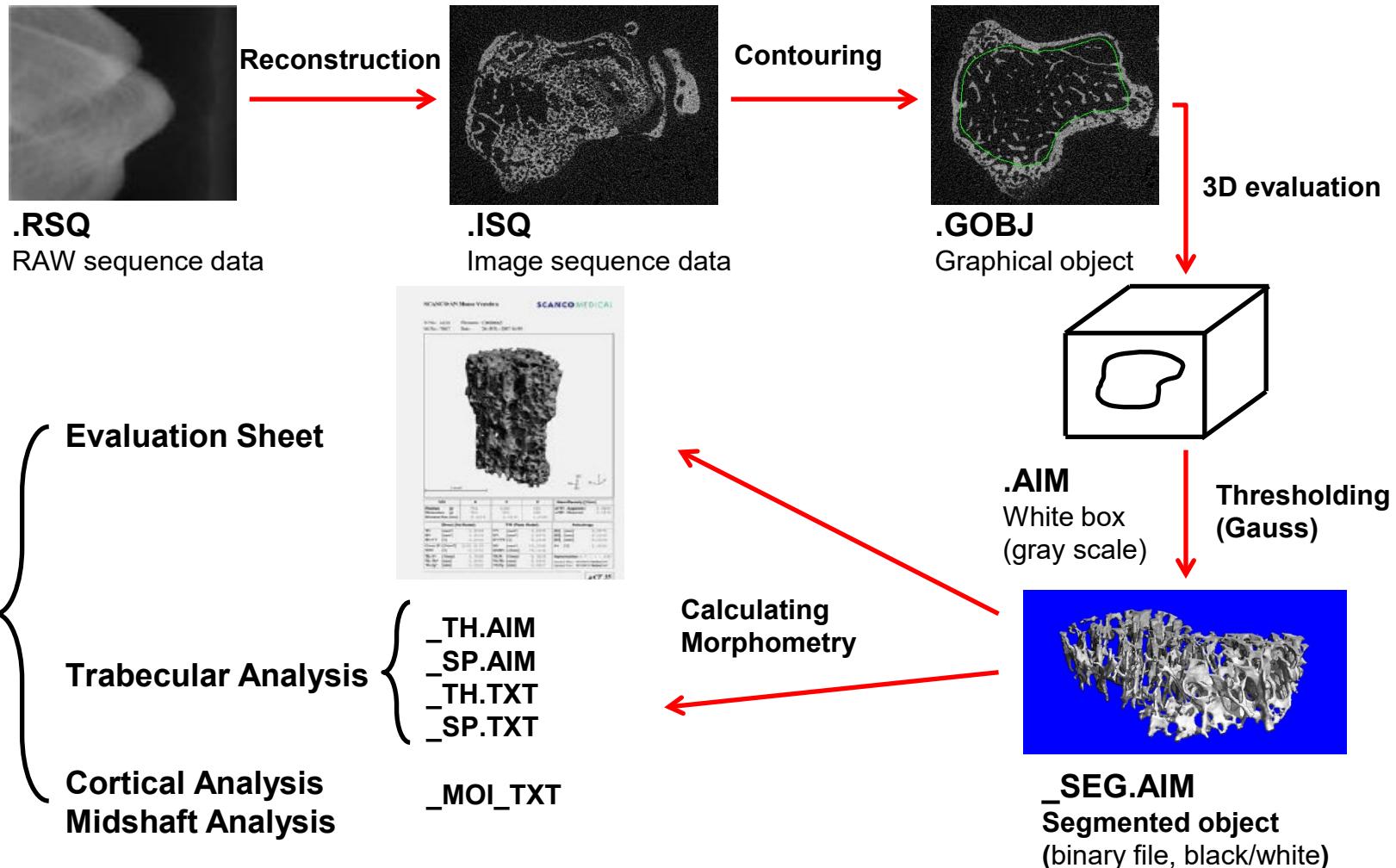
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From Scan to Results

(Trabecular Morphometry, Midshaft Analysis)



How to reduce evaluation time

- **Server computational capacity**
 - 5 evaluation tasks in parallel
- **Pick a lower resolution**
 - Mouse tibia/femur: 7.4 µm (ex vivo), 10 µm (in vivo)
 - Rat tibia/femur: 7.4 µm / 10 µm (ex vivo),
 - Full body skeleton: 50 – 100 µm
 - Imaging only (no quantification): 15 -
- **Use a smaller VOI**
 - Reduce the number of slices
 - Refine ROIs



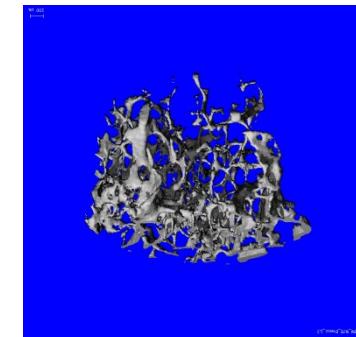
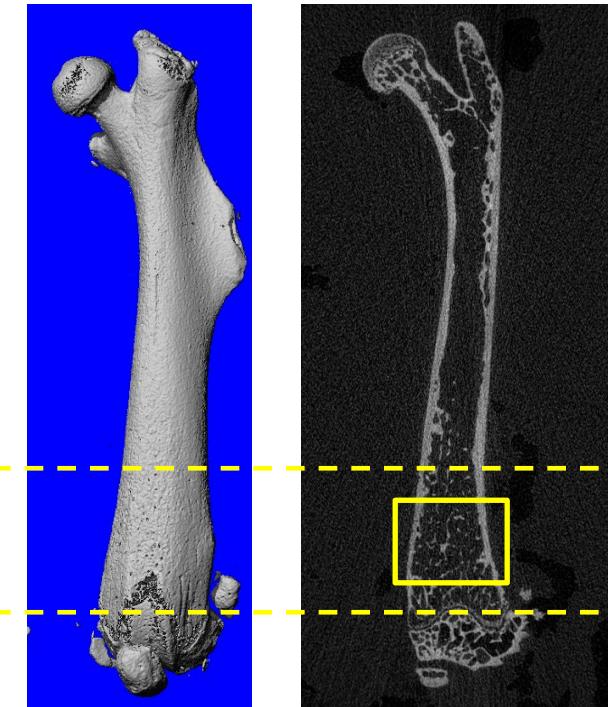
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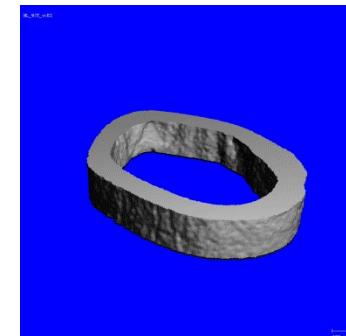
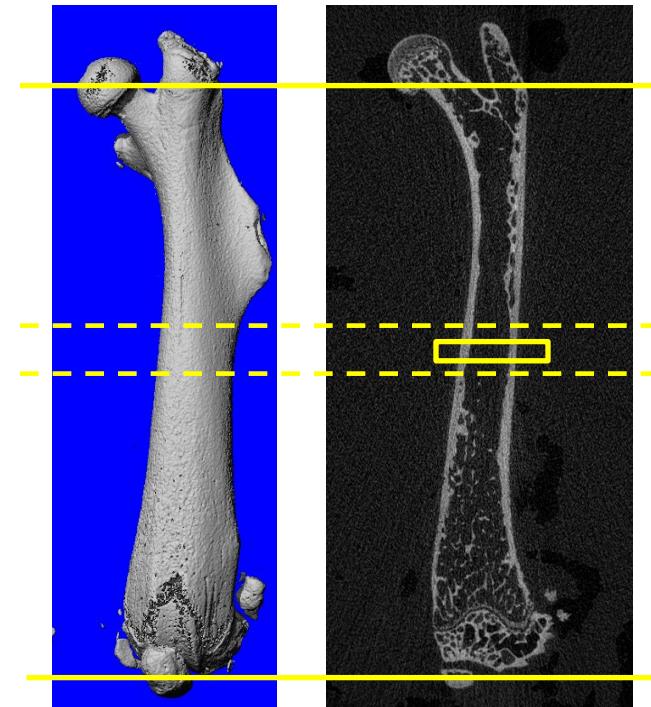
How to reduce evaluation time

- Mouse trabecular bone analysis
 - Distal femur, proximal tibia
 - 6 μm isotropic voxel size
 - Scan Region
 - 1-2 stacks, ~210-420 slices
 - Average scan time: 24-48 mins
 - Analysis region
 - 100-200 slices, 0.5-1mm from the growth plate
 - Outcome measures: BV/TV, Tb.Th, Tb.N, Tb.Sp, SMI, Conn.D, BMD, TMD



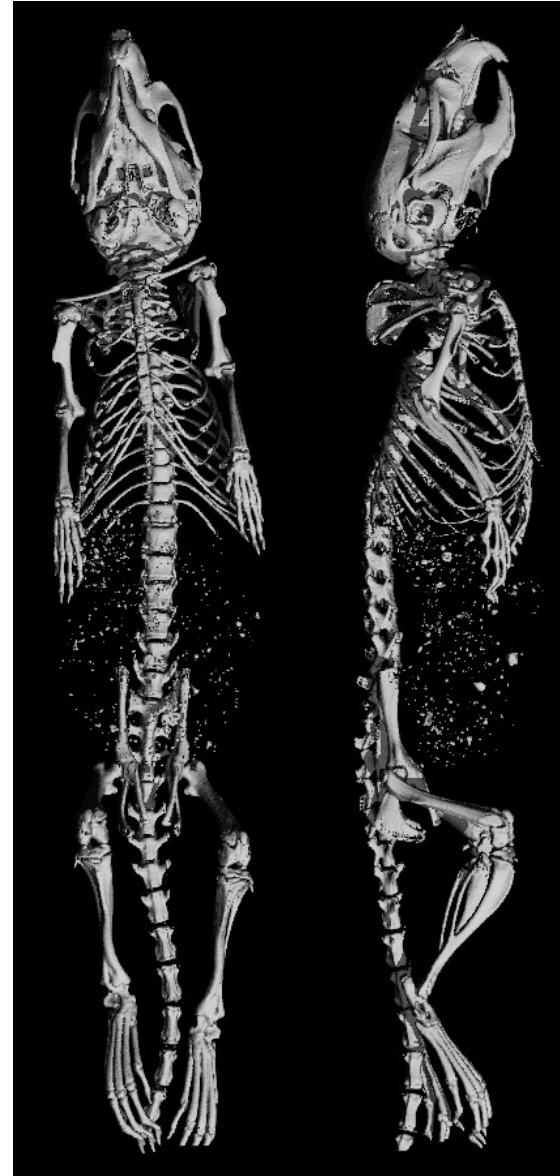
How to reduce evaluation time

- Mouse cortical bone analysis
 - Midshaft of tibia or femur
 - 6-10 μm isotropic voxel size
 - Scan Region
 - 1 stack, ~210 slices
 - Average scan time: 24 mins
 - Analysis region
 - Middle 50 slices
 - Outcome measures: Ct.Area, Ct.Th, pMOI, Ct.Po, TMD



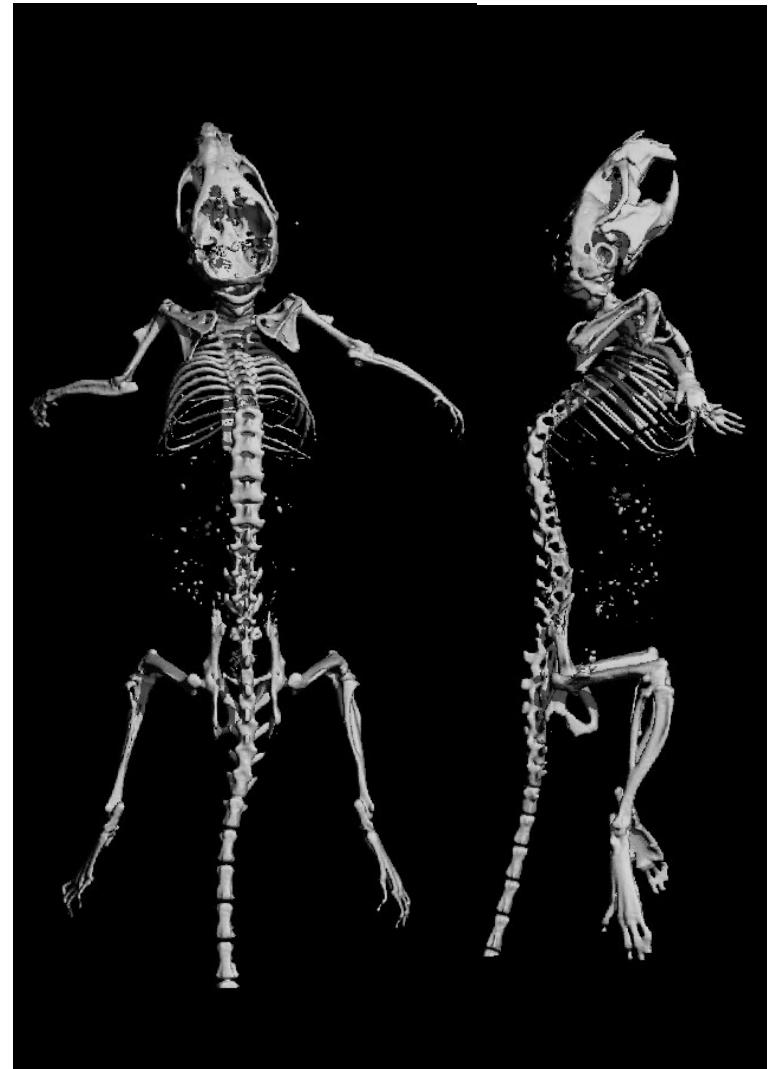
How to reduce evaluation time

- Mouse skeleton imaging
 - Scanner: microCT 45
 - 50 µm isotropic voxel size
 - Scan Region
 - 7-10 stacks, 1500 – 2000 slices
 - Average scan time: 120 mins



How to reduce evaluation time

- Rat skeleton imaging
 - Scanner: vivaCT 80
 - 100 μm isotropic voxel size
 - Scan Region
 - 12 – 13 stacks, 1000 – 1100 slices
 - Average scan time: 25 mins



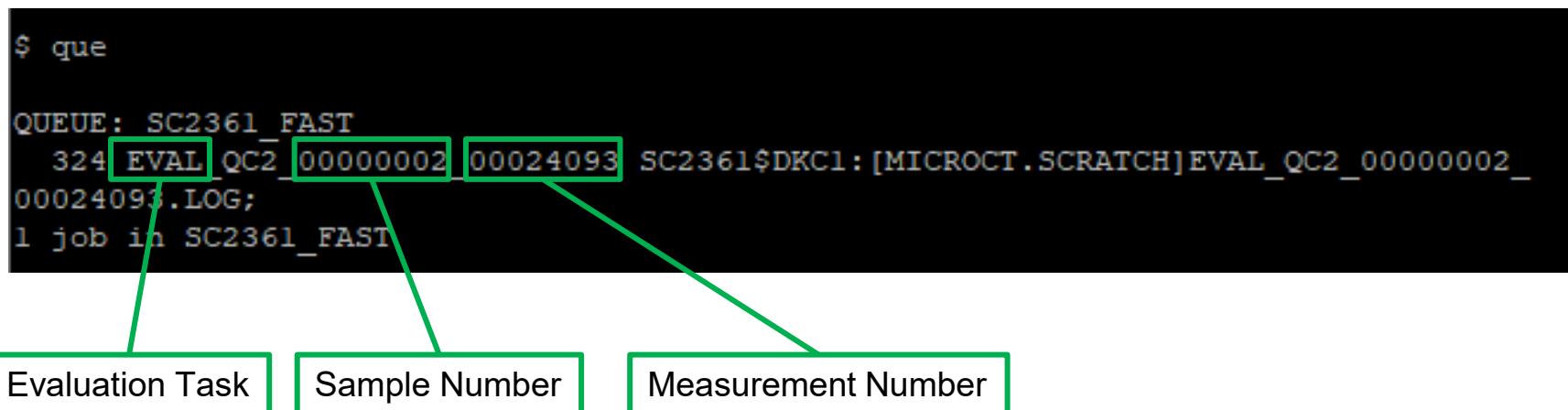
What to do if an evaluation does not run through ?

➤ How to check if an evaluation task completes successfully

- Google Spreadsheet 
- ‘Que’ command
- Check txt / pdf file

```
$ que

QUEUE: SC2361_FAST
 324 EVAL_QC2_00000002_00024093 SC2361$DKC1:[MICROCT.SCRATCH]EVAL_QC2_00000002_
00024093.LOG;
1 job in SC2361_FAST
```



The diagram shows the output of the 'que' command in a terminal window. A green line connects the 'EVAL' in 'EVAL_QC2...' to a box labeled 'Evaluation Task'. Another green line connects the '00000002' to a box labeled 'Sample Number'. A third green line connects the '00024093' to a box labeled 'Measurement Number'.

Evaluation Task Sample Number Measurement Number



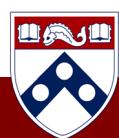
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(Trabecular Morphometry)

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Support Threshold	Unit	Data-Threshold	VOI	X	Y	Z	Mean/Density [mg HA/ccm]
1.20	2	140	Position [p]	1506	1029	704	of TV (Apparent) -24.4381
			Dimension [p]	820	752	464	of BV (Material) 502.2062
			Element Size [mm]	0.0030	0.0030	0.0030	
a-Threshold	VOX-TV	VOX-BV	VOX-BV/TV	Conn	Direct (No Model)	TRI (Plate Model)	Anisotropy
140	6	4587.000			TV [mm ³] 6.0222	TV [mm ³] -	H1 [mm] -
					BV [mm ³] 0.0480	BV [mm ³] -	H2 [mm] -
					BV/TV [l] 0.0080	BV/TV [l] -	H3 [mm] -
Conn-Dens.	TRI-SMI	DT-Tb.N	DT-Tb.Th		Conn. D. [1/mm ³] -	BS [mm ²] -	DA [l] -
6.0222	0.0480	0.0080			SMI [l] -	BS/BV [1/mm] -	
TRI-SMI	DT-Tb.N	DT-Tb.Th	DT-Tb.Sp		Tb.N* [1/mm] -	Tb.N [1/mm] -	Segmentation: 1.2 / 2 / 140
0.0480	0.0080	-	-		Tb.Th* [mm] -	Tb.Th [mm] -	Operator Meas.: PCMD MicroCT Imaging
					Tb.Sp* [mm] -	Tb.Sp [mm] -	Operator Eval.:

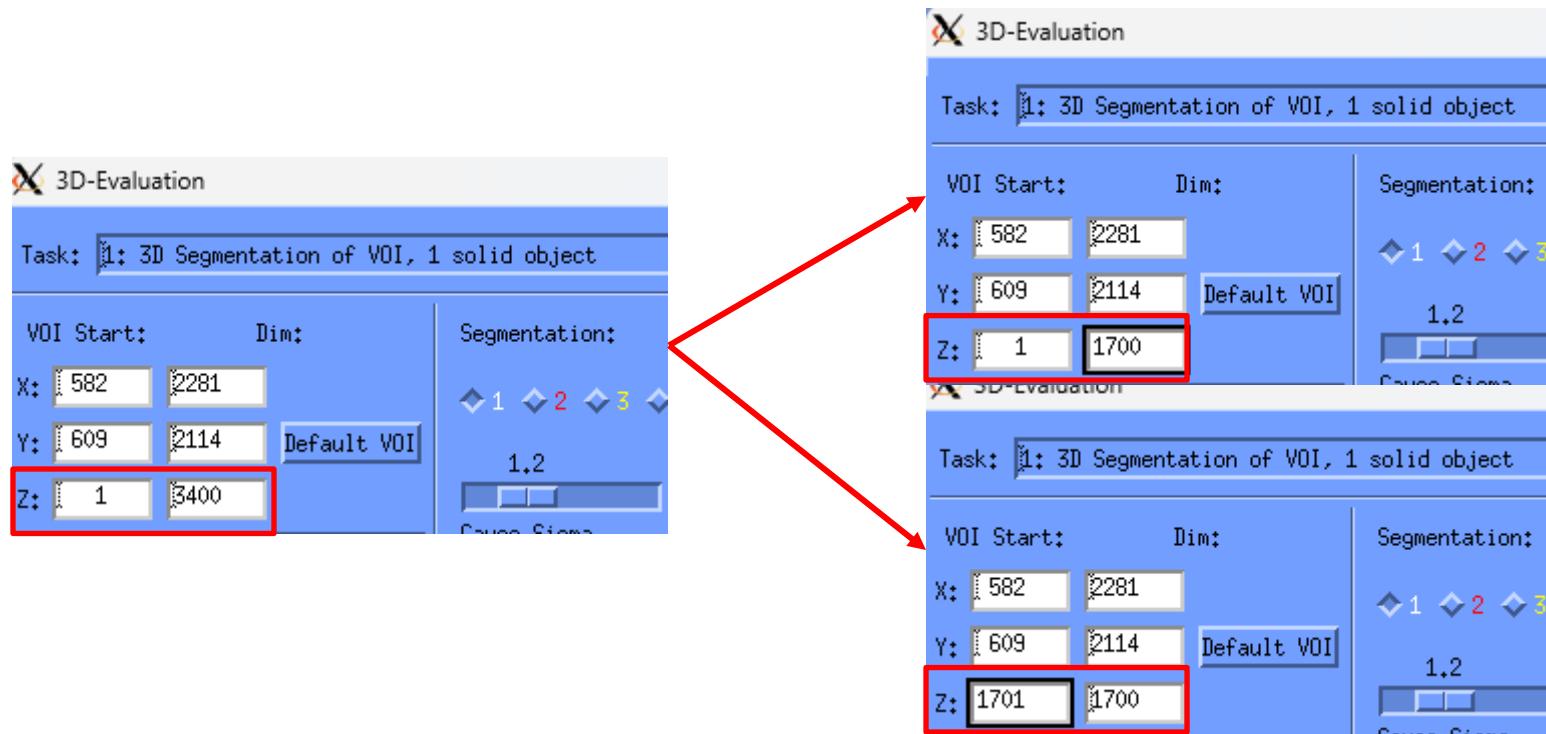


What to do if an evaluation does not run through ?

(Trabecular Morphometry)

➤ Task break up

- Slice number < 3000
- Statistics – combine the results by taking average
- 3D image – align multiple images into a full view



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